



BURMESE PYTHON RANGE DELIMITATION USING ENVIRONMENTAL DNA SURVEYS IN THE GREATER EVERGLADES ECOSYSTEM

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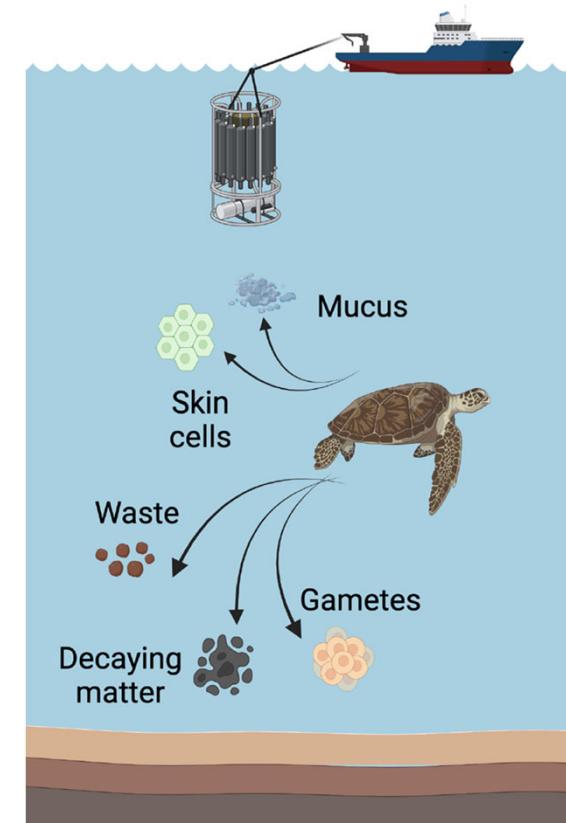
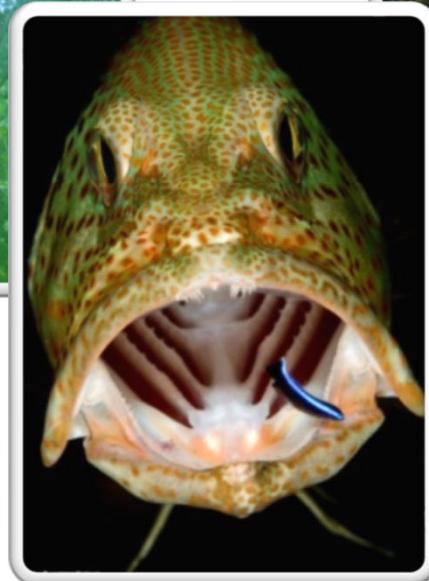
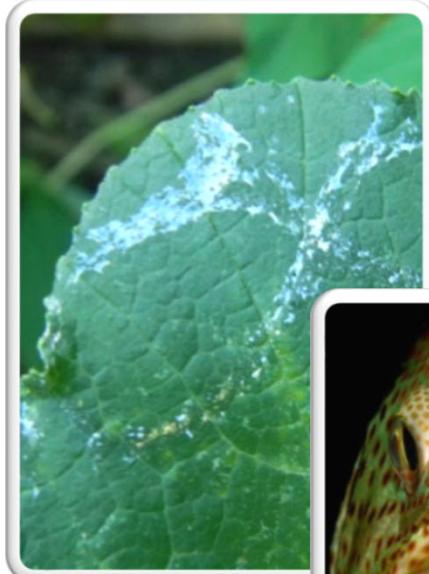
*USGS Wetland and Aquatic Research Center
WARC Conservation Genetics Laboratory*

*This project is funded by USGS's Greater Everglades
Priority Ecosystems Science (GEPES) Program*

U.S. Department of the Interior
U.S. Geological Survey

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ENVIRONMENTAL DNA SOURCES



DETECTION OF EDNA MOLECULES, NOT THE SPECIES ITSELF

SUSIE THEROUX, SCCWRP

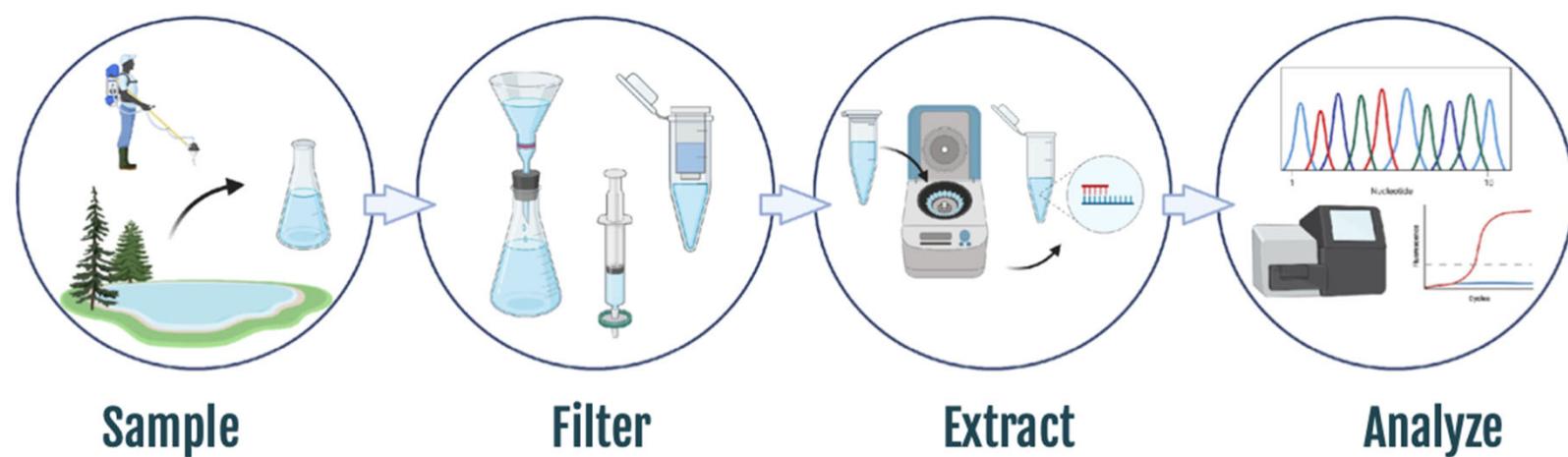
ENVIRONMENTAL DNA METHODS

- Non-invasive sampling
- Sensitive and specific
- High catch-per-unit



* Stock Imagery

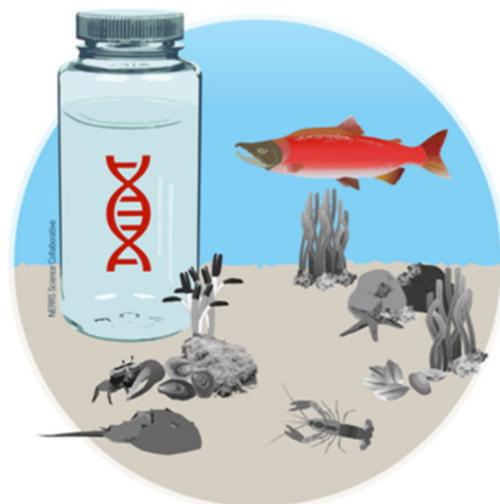
eDNA ANALYSIS PROCESS



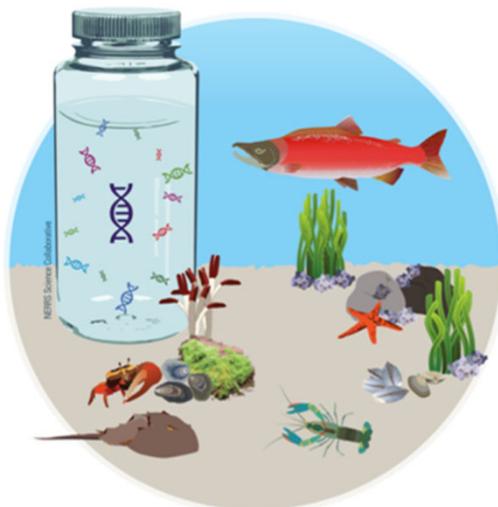
SUSIE THEROUX, SCCWRP

TWO PRIMARY eDNA DETECTION METHODS

Targeted:
Single species or genera

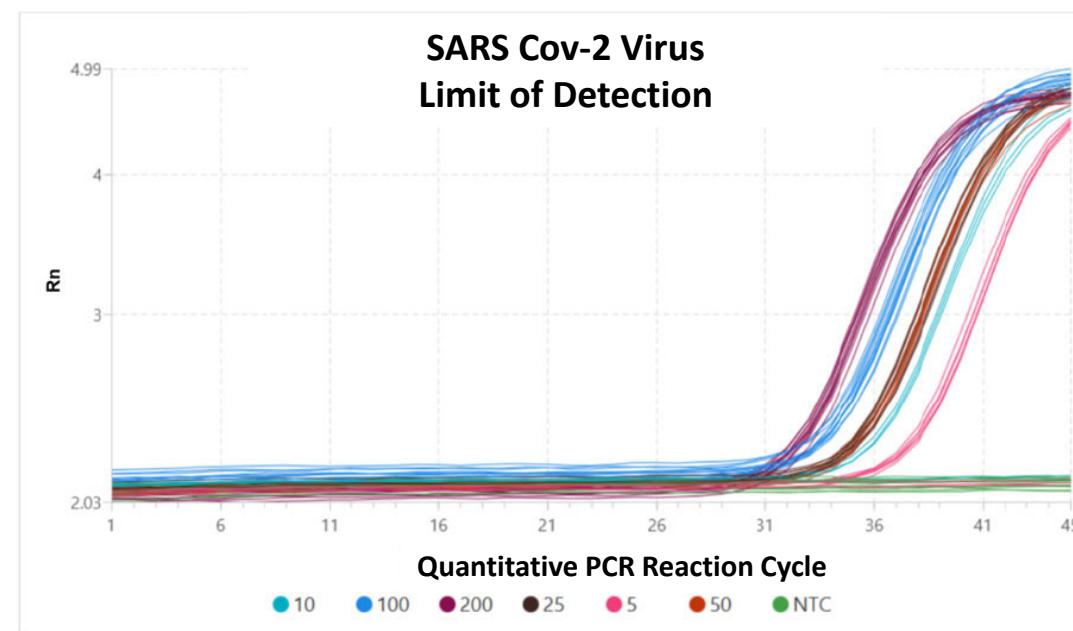
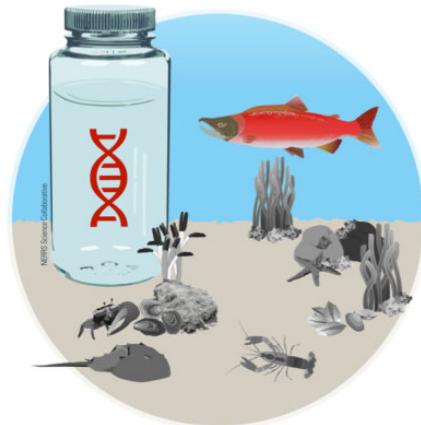


Metabarcoding: Community assessment

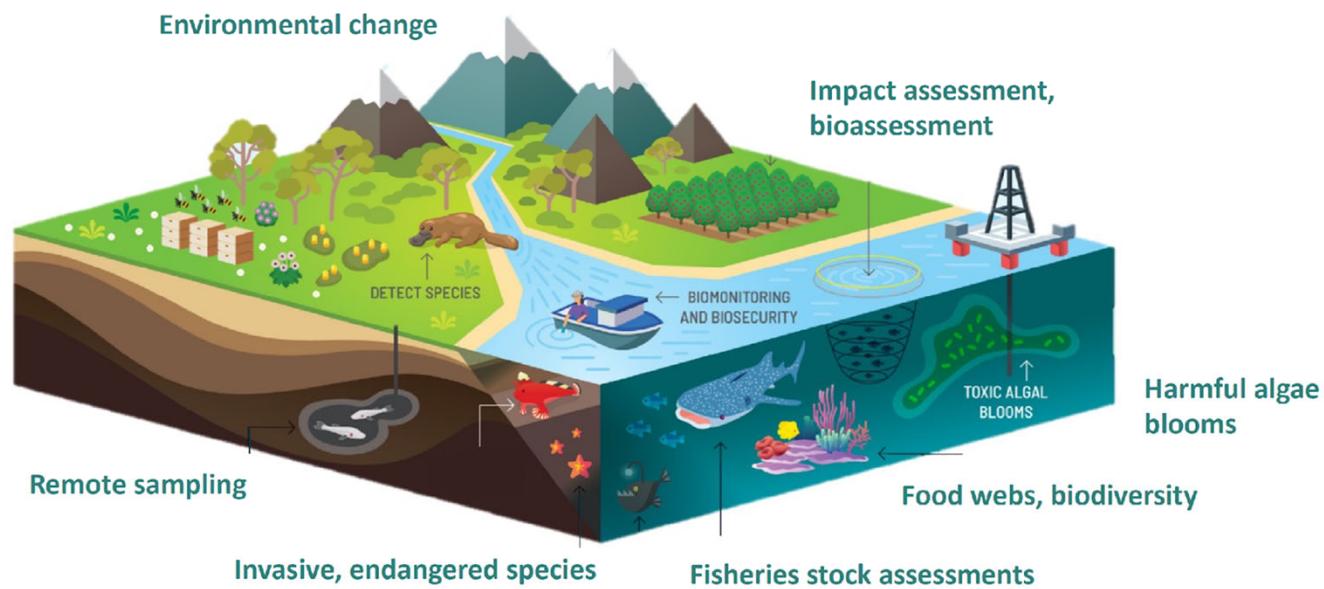


Targeted eDNA

- Species-specific
- Sensitive
- Detection/non-detection of eDNA
- Occurrence and detection rates



eDNA applications



Modified from Berry et al., 2020 Environmental DNA





INVASIVE SPECIES eDNA APPLICATIONS

- Inform EDRR efforts
- Monitor control or eradication measures
- Inform invasion fronts, pathways or hot spot areas
- Occupancy estimates
- Food web analyses

IMPERILED SPECIES eDNA APPLICATIONS

- Species range/habitat delimitation
- Travel corridors
- Monitor recovery efforts (increasing occupancy)
- Assist with extinction determinations





U.S. NATIONAL AQUATIC eDNA STRATEGY

The White House Office of Science and Technology Policy (OSTP)

National Science and Technology Council (NSTC)
Biodiversity Working Group, eDNA Task Team

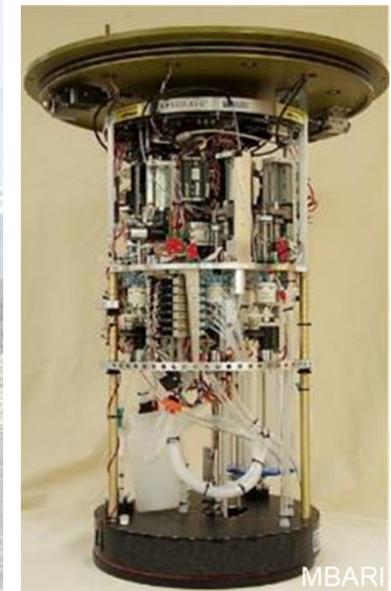
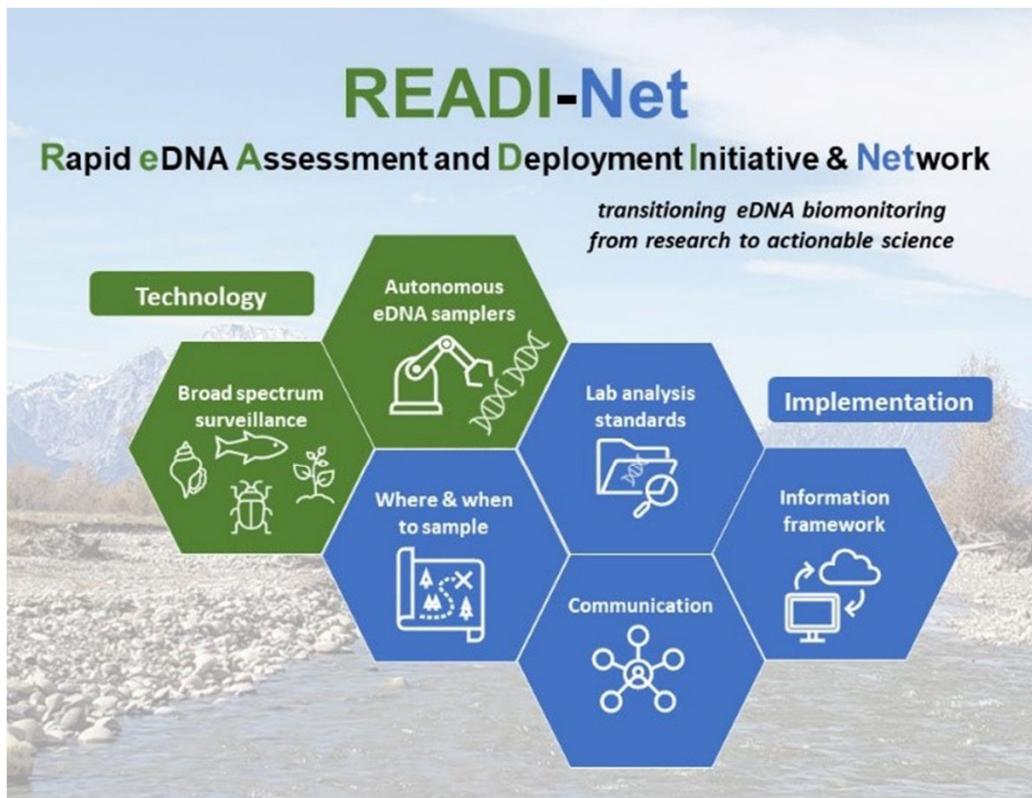
*Harnessing the power of eDNA to explore, map, monitor,
and better understand aquatic life to sustain and restore
biological resources into the future*



NATIONAL eDNA ACTIVITIES



- Bipartisan Infrastructure Law Funds
- Early Detection Rapid Response (EDRR) National Framework
 - Molecular Lab Network
 - NAS database eDNA reporting
 - READI-Net (robotic sampling and data standards)



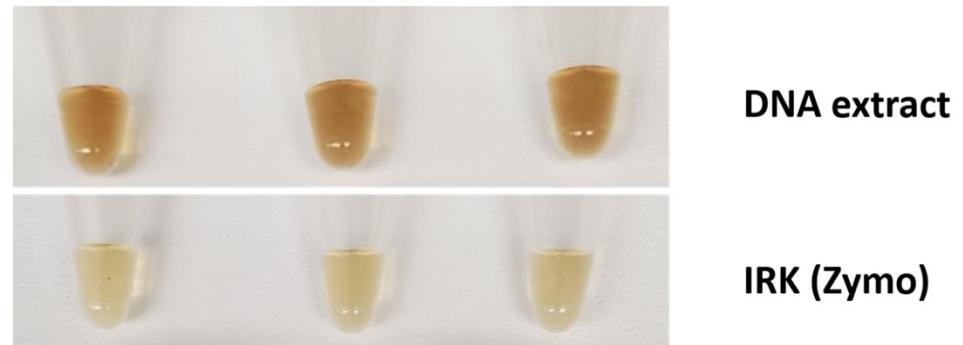
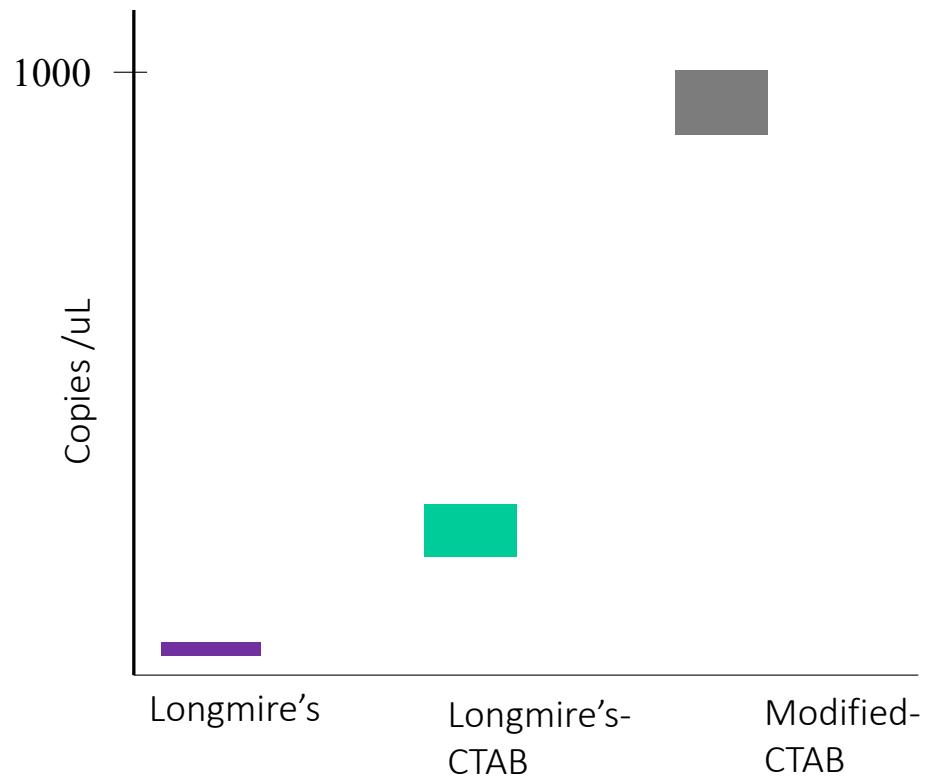
- Adam Sepulveda, USGS

SOUTHEAST eDNA STUDIES



Descriptions in Hunter et al. (2019)

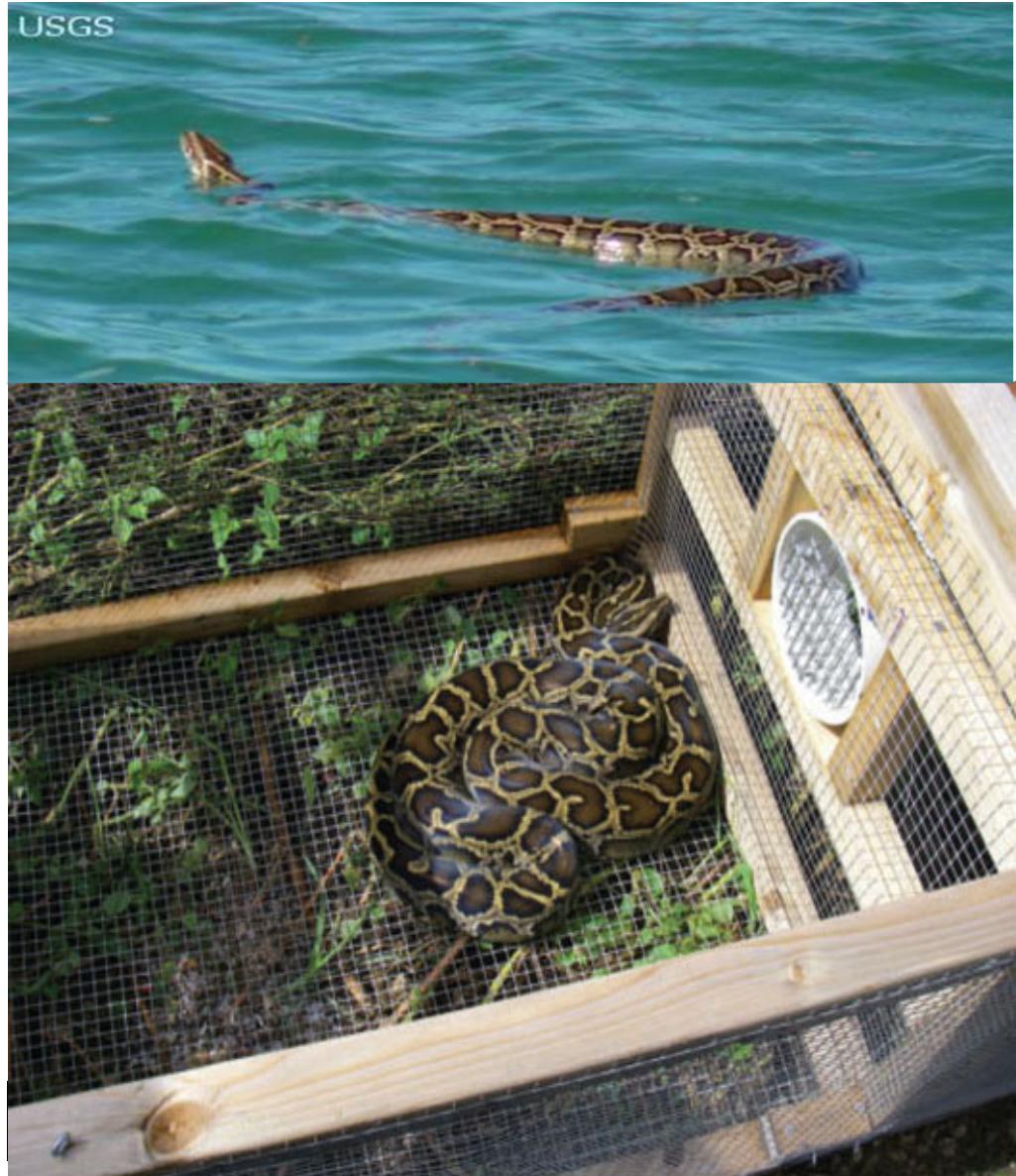
SOUTHEAST eDNA STUDIES: PCR Inhibitors



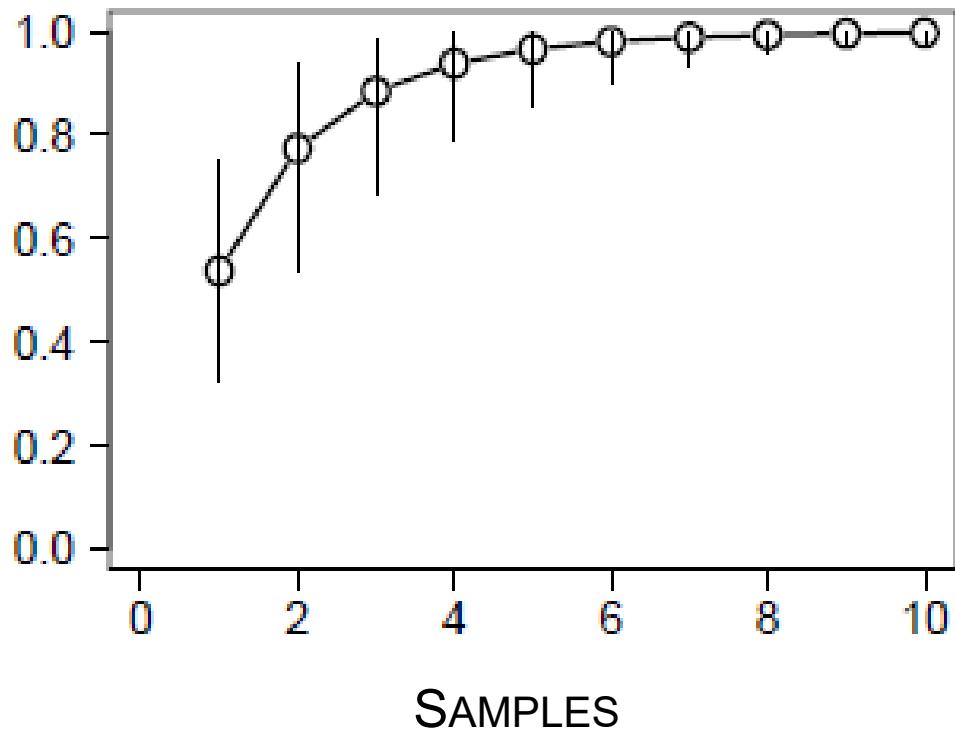
DESCRIPTIONS IN HUNTER ET AL. (2019)

Invasive Burmese python

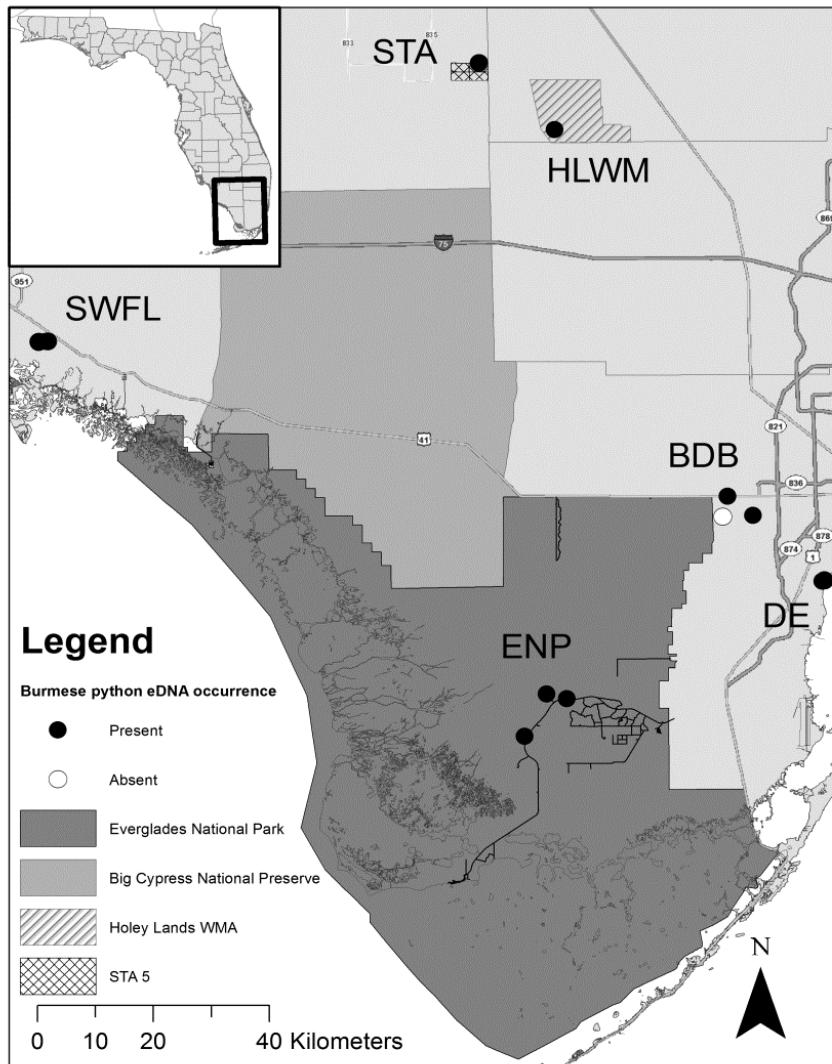
- Hindering restoration efforts
- Semi-aquatic, cryptic
- Visual sighting and Trapping rate is < 1%



- Visual sighting and trapping capture < 1%
- eDNA detection is <40-100%
 - Occupancy analyses
 - Probability of occurrence



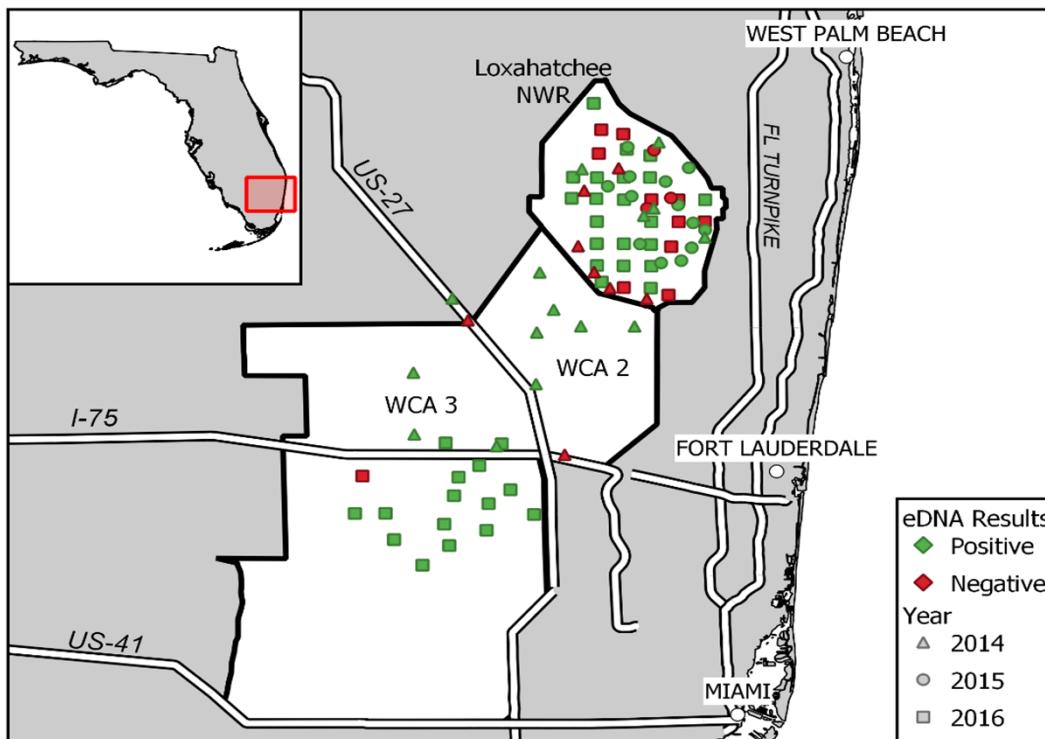
BURMESE PYTHON eDNA DETECTIONS: 37 OF 63 SAMPLES



HUNTER ET AL., PLOS ONE 2015

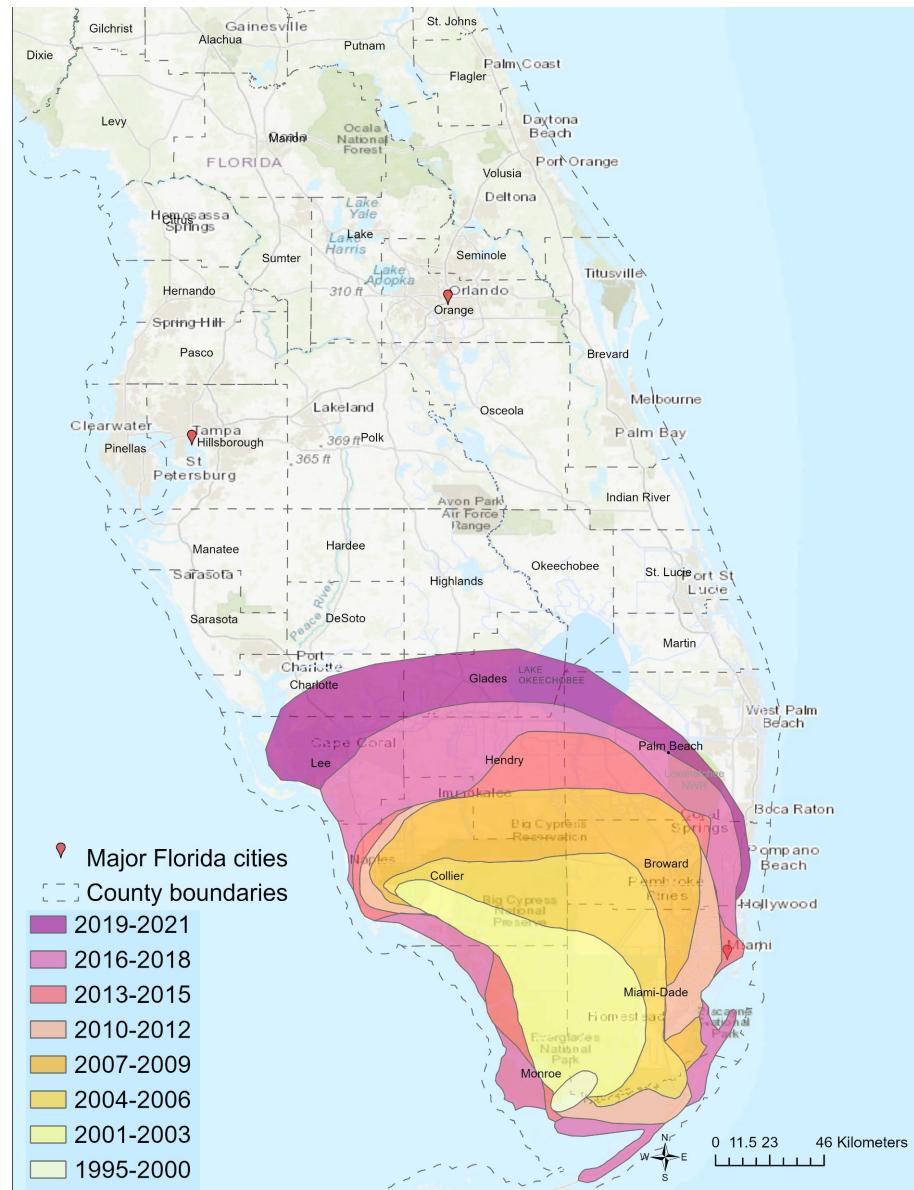


eDNA detections in Loxahatchee National Wildlife Refuge



HUNTER ET AL. (2019) ECOLOGICAL INDICATORS

CURRENT ESTIMATION OF INVASION FRONT

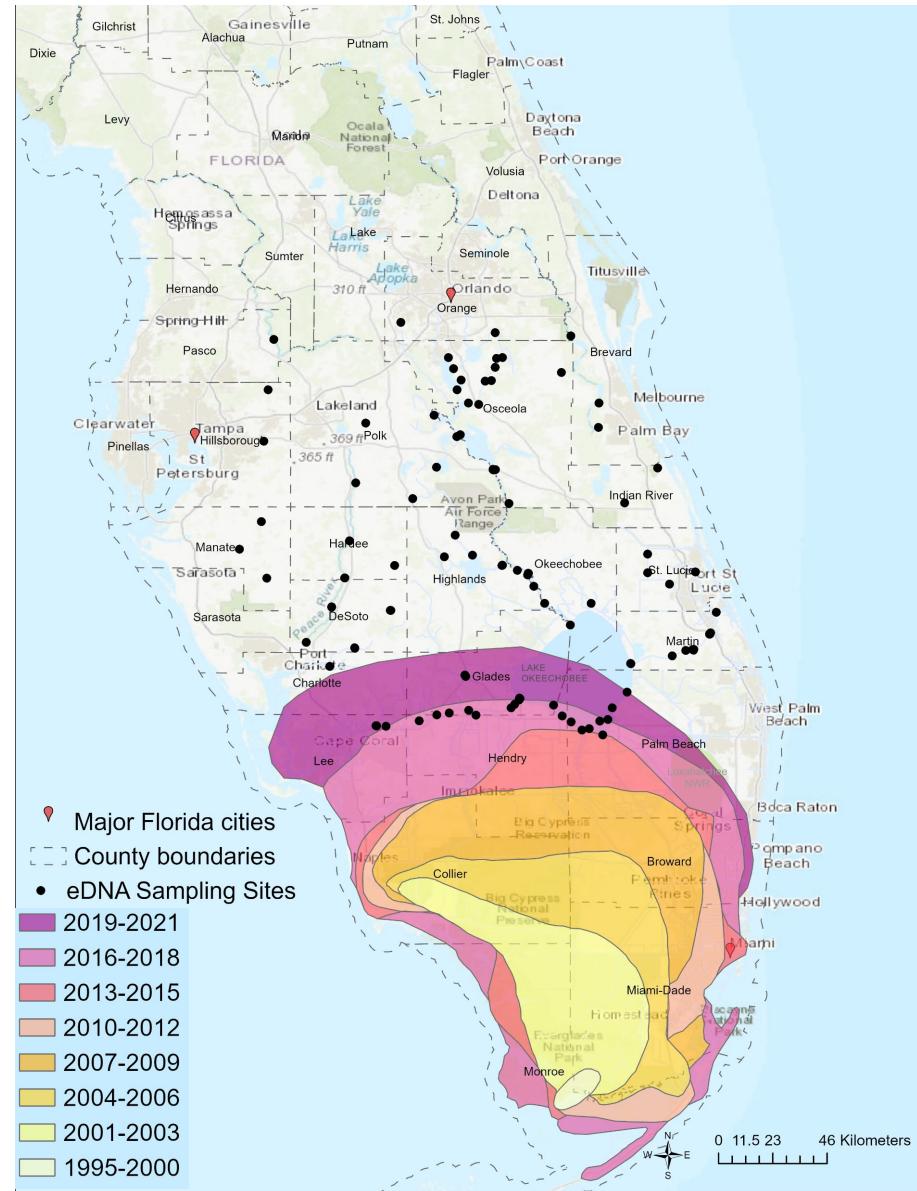


Polygons from Guzy et al. (2023)



TOTAL PROJECT EFFORT

Preliminary Information-Subject to
Revision. Not for Citation or
Distribution.



2019-2023
Total sampled sites = 141
Sites resampled = 26

Polygons from Guzy et al. (2023)

ALL SAMPLES

2019-2023

Preliminary Information-Subject to
Revision. Not for Citation or
Distribution.



- POSITIVE DETECTION
- MIXED DETECTION
- NO DETECTION

DISCUSSION

- Likely detecting a population up to south Orlando
 - Not all recent releases
- The population may extend beyond our detections
- Evidence for more detections on the western side of the peninsula
 - Possibly indicating a corridor north



Priority areas for further sampling?

Preliminary Information-Subject to
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THANK YOU!

- Special thanks to John Stechly, Alexis Tays, Dr. Jason Ferrante, Stephanie Katircioglu, Dr. Maite de Maria, Maggie Parrish, Garet Dicks, Chelicia Espinoza, India Sander-Nazario for all the help in the lab and field.
- *We are grateful for the support by USGS's Greater Everglades Priority Ecosystems Science (GEPES) Program*



REFERENCES

- Guzy, J. C., Falk, B. G., Smith, B. J., Willson, J. D., Reed, R. N., Aumen, N. G., ... & Hart, K. M. (2023). Burmese pythons in Florida: A synthesis of biology, impacts, and management tools. *NeoBiota* 80: 1–119.
- Hunter, M. E., Oyler-McCance, S. J., Dorazio, R. M., Fike, J. A., Smith, B. J., Hunter, C. T., Reed, R.N. & Hart, K. M. (2015). Environmental DNA (eDNA) sampling improves occurrence and detection estimates of invasive Burmese pythons. *PloS one*, 10(4), e0121655.
- Hunter, M. E., Ferrante, J. A., Meigs-Friend, G., & Ulmer, A. (2019a). Improving eDNA yield and inhibitor reduction through increased water volumes and multi-filter isolation techniques. *Scientific Reports*, 9(1), 5259.
- Hunter, M. E., Meigs-Friend, G., Ferrante, J. A., Smith, B. J., & Hart, K. M. (2019b). Efficacy of eDNA as an early detection indicator for Burmese pythons in the ARM Loxahatchee National Wildlife Refuge in the greater Everglades ecosystem. *Ecological Indicators*, 102, 617-622.



2022 EDNA RESULTS AND EDDMAPS DETECTIONS

■ EddMap detection

● eDNA no detection

○ eDNA detection

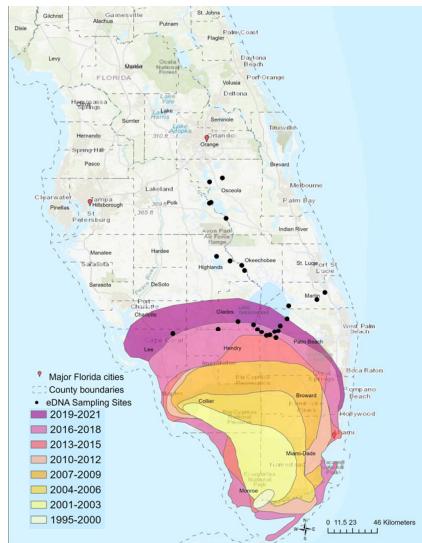
Areas of interest



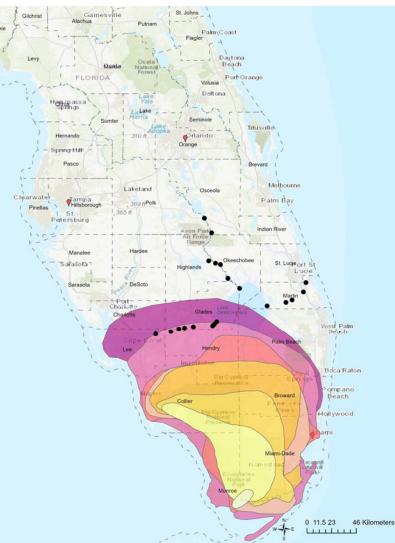
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SAMPLING DESIGN BY YEAR

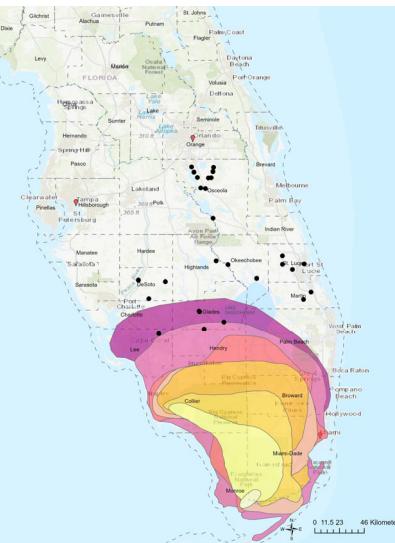
Fall 2019



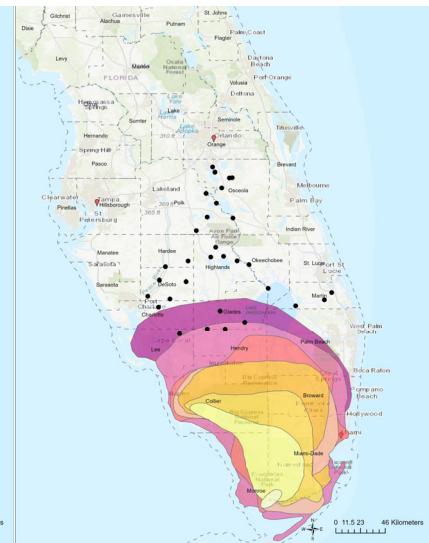
Fall 2020



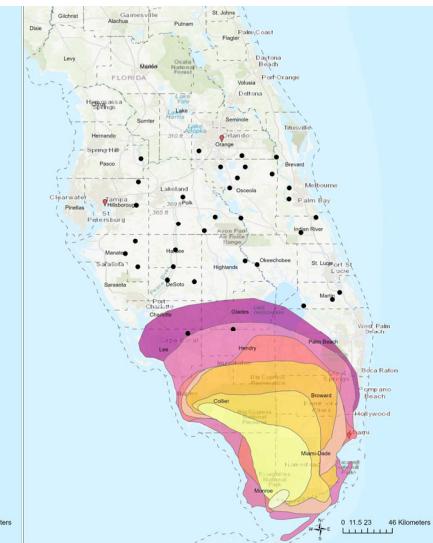
Spring/Fall 2021



Spring 2022



Spring 2023



N = 26

N = 20

N = 28

N = 31

N = 34



PRELIMINARY INFORMATION-SUBJECT TO REVISION. NOT FOR CITATION OR DISTRIBUTION.

CHALLENGES WITH eDNA METHODS

- eDNA METHODS DO NOT DETECT THE SPECIES ITSELF
- MULTIPHASE FIELD AND LAB PROCESS, SENSITIVE DATA INTERPRETATIONS
- EACH SPECIES AND HABITAT REQUIRES UNIQUE METHODS



ALL SAMPLE SITES AND DETECTIONS

- POSITIVE DETECTION
- MIXED DETECTION
- NO DETECTION

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DETECTIONS BY YEAR

POSITIVE DETECTION MIXED DETECTION NO DETECTION

2019

2020

2021

2022

2023

N = 26

N = 20

N = 28

N = 31

N = 34



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